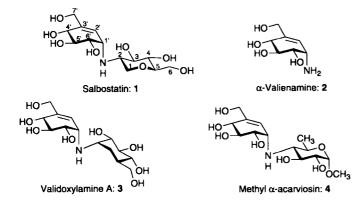
Total Synthesis of the Trehalase Inhibitor Salbostatin

Tatsuya Yamagishi, Chikara Uchida, and Seiichiro Ogawa*

Abstract: The total synthesis of trehalase inhibitor salbostatin (1), recently discovered as a novel metabolite of *Streptomyces albus* species, has been achieved starting from the major product (58% yield) from the coupling of the amine di-O-isopropylidene- α -valienamine and the electrophile 1,5:2,3-dianhydro-D-mannitol in 2-propanol. Deprotection with aqueous acetic acid and subsequent purification on a column of acidic resin afforded 1, which showed inhibitory activity (IC₅₀ = 8.3 μ M L⁻¹) against silkworm trehalase.

Introduction

Very recently, new trehalase inhibitor salbostatin (1) was discovered^[1] as a novel metabolite of *Streptomyces albus*, ATCC 21838, and the structure has been established mainly on the basis of ¹H NMR spectroscopic data.^[2] Salbostatin inhibits



trehalase from porcine kidneys with an inhibition constant K_i of 1.8×10^{-7} M, and possesses a very unique pseudo-disaccharide structure composed of 2-amino-1,5-anhydro-2-deoxy-D-glucitol attached by way of an imino bridge to an unsaturated 5a-carba-sugar,^[**] the α -valienamine (2) residue. Similar pseudo-disaccharidic glycosidase inhibitors containing 2, namely, validoxylamine A (3)^[3] and methyl acarviosin (4),^[4] a core component of acarbose, are known. The former is a potent trehalase inhibitor and its dihydro derivative,^[3] most probably mimicking the

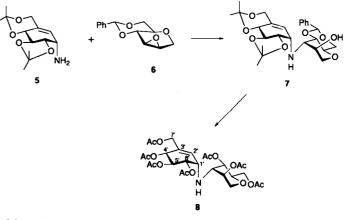
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 α, α -trehalose structure, also exhibits high inhibitory activity. The latter is a strong α -glucosidase inhibitor. In both cases, the valienamine portions are thought to be mimics of the glucopyranosyl cations proposed to be formed during hydrolysis of gly-cosides.

We have been studying the relationship between structure and inhibitory activity in this class of compound, ^{[51} and our interests are now especially focused on trehalase inhibitors such as 3 and trehazolin.^[3, 6] This paper describes a convenient total synthesis of salbostatin, which serves both to confirm the proposed structure and to elaborate a general method for the preparation of salbostatin analogues. The method involves coupling of an anhydro sugar with the versatile unsaturated carbasugar donor 2,3:4,6-di-*O*-isopropylidene- α -valienamine (5).^[7]

Results and Discussion

First, coupling of the amine 5 and readily accessible 1,5:2,3-dianhydro-4,6-O-benzylidene-D-mannitol (6)^[8] (1.3 molequiv) was carried out in 2-propanol in a sealed tube for eight days at 120 °C (Scheme 1). The reaction was very slow and, after com-



Scheme 1.

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^[**] In order to differentiate it from the exo-methylene derivative with unsaturation between C-5 and C-6, we have named the unsaturated carbasugar "α-valienamine", which has unsaturation located between C-5 and C-5a, 5a-carba-α-Dxylo-hex-5(5a)-enopyranosylamine.

plete disappearance of 5, the product 7, contaminated with some deprotected derivatives, was hydrolyzed with aqueous acetic acid and acetylated. A single hepta-N, O-acetyl derivative 8 was isolated (89% yield). The opening of the epoxide proceeded with high regioselectivity. The ¹H NMR spectrum of 8 was, however, different from that reported for an authentic sample of peracetylated 1,^[2] and was consistent with the expected product resulting from *trans*-diaxial opening of the epoxide 6. Thus, the ¹H NMR spectrum contained three coupled doublets of doublets at $\delta = 4.73$ (J = 1.5 and 3.7 Hz), 3.32 (J = 3.7 and 4.4 Hz), and 5.09 (J = 4.4 and 10.3 Hz) corresponding to protons at positions 2, 3, and 4, respectively. Regioselective attack of the amine 5 at C-3 seems to be facilitated not only by steric factors, but also by hydrogen bonding between an acetal oxygen atom and the incoming amino function in the transition state.

We therefore chose to use the unprotected 1,5:2,3-dianhydro-D-mannitol (9) as electrophile, which was derived by hydrogenolysis (10% Pd/C) of 6 (95% yield). Removal of the benzylidene group from 6 seemed to enhance the reactivity of the 2,3-epoxide, and, it was hoped, that steric factors would now favor the desired selectivity of the ring opening. Coupling of 5 and a slight excess of 9 (1.3 molequiv) was carried out in 2propanol at 120 °C (Scheme 2). After four days, TLC clearly

showed that two coupling products had been formed. Chromatography of the mixture on a silica gel column with butanone/toluene (3:1, v/v) as eluent afforded the diequatorially opened product 10 (58%) and the diaxially opened product 11 (25%). The structures of 10 and 11 were tentatively assigned as depicted in Scheme 2 on the basis of the ¹H NMR spectra. The spectrum of 10 (270 MHz, CDCl₃) was well resolved and was amenable to a full analysis. Thus, the coupled signals due to 1,1-H and 2-H appear as a doublet of doublets ($\delta = 4.08$, J =4.8 and 11.0 Hz), a triplet ($\delta = 3.19$, J = 11.0 Hz), and a doublet of doublets of doublets ($\delta = 2.80$, J = 4.8, 9.2, and 11.0 Hz), respectively; this is consistent with a 1,5-anhydroglucitol structure of the sugar moiety. The spectrum of 11, on the other hand, consisted of complex multiplets for the ring protons of the sugar fragment. Preferential attack of the amine at C-2 of 9 might be partly rationalized, in addition to stereoelectronic effects, by assuming that 9 adopts the flipped ${}^{1}C_{4}$ conformation for trans-diaxial cleavage, favored by hydrogen bonding between the epoxide oxygen atom and the 6-hydroxyl group. O-Deisopropylidenation of 10 was achieved in aqueous 70% acetic acid for one hour at 60 °C to give, after purification on a column of Dowex 50 W-X2 (H⁺) resin with 1 M aqueous ammonia, salbostatin 1 in 97% yield. The ¹³C and ¹H NMR spectra were in good agreement with those of an authentic sample.^[2] To further confirm its identity, the synthetic sample of 1 was converted to the hepta-O-acetyl derivative 12, the ¹H NMR spectrum of which was shown to be superimposable on that reported.^[2] Biological assay demonstrated that 1 possesses moderate inhibitory activity $(IC_{50} = 8.3 \times 10^{-6} \text{ M})$ against silkworm trehalase, rather lower than that of validoxylamine A^[3] and trehazolin.^[6]

The first total synthesis of the inhibitor salbostatin 1, presented here, confirms the structure proposed previously, and also provides a convenient method for preparation of its analogues.

Experimental Procedure

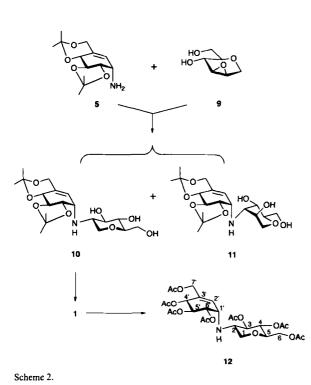
General method: Specific rotations: Jasco DIP-370 digital polarimeter in 1 dm cells. IR spectra: Jasco IR-810 spectrometer. ¹³C and ¹H NMR spectry: Jeol JNM GSX-270 FT (270 MHz) instrument. TLC: silica gel 60 GF (Merck); detection by charring with conc. sulfuric acid. Column chromatrography: silica gel Wakogel C-300 (Wako, Osaka). Solvents: after drying them with anhydrous sodium sulfate, solvents were evaporated below 50 °C under reduced pressure.

2,4,6-Tri-O-acetyl-1,5-anhydro-3-deoxy-3-[2,3,4,6-tetra-O-acetyl-5a-carba-xylo-

hex-5(5a)-enopyranosylamino|-D-altritol (8): A solution of 2,3:4,6-di-O-isopropylidene-a-valienamine [7] (5, 15.5 mg, 0.061 mmol) and 4,6-O-benzylidene-1,5:2,3-dianhydro-D-mannitol [8] (6, 18.5 mg, 0.079 mmol, 1.3 molequiv) in 2-propanol (0.3 mL) was heated in a sealed tube for 8 d at 120 °C and then evaporated. The residue was treated with 70% aq. acetic acid (2 mL) for 3 h at 70°C. The mixture was evaporated and the residue was eluted from a column of silica gel (2 g) with acetone/toluene (1:2, v/v) as eluent to give a crude product, which was acetylated with acetic anhydride and pyridine. Chromatography on Dowex 50 W-X2 (H⁺) resin (2 mL) with 1 M aq. NH, OH gave a single hepta-N, O-acetyl derivative 8 (33.3 mg, 89%) as a syrup; $[\alpha]_{D}^{22} = +78$ (c = 1.4 in CHCl₃); ¹H NMR (270 MHz, CDCl₃, TMS): $\delta = 5.98$ (1 H, d, J(1',5'a) = 5.1 Hz, 5'a-H), 5.50 (1 H, t, J(2',3') = J(3',4') = 6.6 Hz, 3'-H), 5.46 (1 H, d, J(3',4') = 6.6 Hz, 4'-H), 5.09 (1 H, dd, J(3,4) = 4.4, J(4,5) = 10.3 Hz, 4-H), 5.02 (1 H, dd, J(1',2') = 4.8, J(2',3') =6.6 Hz, 2'-H), 4.73 (1 H, dd, J(1,2) = 1.5, J(2,3) = 3.7 Hz, 2-H), 4.64 and 4.37 (each 1 H, ABq, J(gem) = 13.2 Hz, 6',6'-H), 4.22 (1 H, dd, J(5.6a) = 5.5, J(gem) = 5.512.1 Hz, 6a-H), 4.15 (1 H, dd, J(5,6b) = 4.0, J(gem) = 12.1 Hz, 6b-H), 4.11 (1 H, dd, J(1,2) = 1.5, J(gem) = 12.5 Hz, 1-H), 3.92 (1 H, ddd, J(5,6a) = 5.5, J(5,6b) = 12.5 Hz, 1-H), 3.92 (1 H, ddd, J(5,6a) = 5.5 4.0, J(4,5) = 10.3 Hz, 5-H), 3.83 (1 H, d, $J(1,2) \approx 0$, J(gem) = 12.5 Hz, 1-H), 3.53 (1 H, dd, J(1',2') = 4.8, J(1',5'a) = 5.1 Hz, 1'-H), 3.32 (1 H, dd, J(2,3) = 3.7,J(3,4) = 4.4 Hz, 3-H), 2.13, 2.11, 2.10, 2.08, 2.07, 2.063, 2.056 (each 3H, 7s, $7 \times Ac$; the signal due to NH proton was not detected; $C_{27}H_{37}NO_{15}$ (615.6): calcd C 52.68, H 6.06, N 2.28; found C 52.51, H 6.17, N 2.37.

1,5:2,3-Dianhydro-D-mannitol (9): A solution of 4,6-*O*-benzylidene-1,5:2,3-dianhydro-D-mannitol (6, 134 mg, 0.57 mmol) in ethanol (2 mL) was hydrogenolyzed in the presence of 10% Pd/C (10 mg) under an atmosphere of hydrogen for 18 h at room temperature. The catalyst was removed by filtration, and the filtrate evaporate d. Chromatography of the residue on a silica gel column (4 g) with ethanol/toluene (1:8, v/v) as eluent to give **9** (79 mg, 95%) as needles, m.p. 111-114 °C (from ethanol), $[\alpha]_D^{25} = +19$ (c = 0.90 in H₂O); ¹H NMR (270 MHz, D₂O, acetone as ref.): $\delta = 4.03$, 3.79 (2d, J(1 gem) = 13.7 Hz, each 1H, 1,1-H), 3.69 (dd, J(5,6) = 2.4, J(6 gem) = 12.3 Hz, 1H, 6-H), 3.51 (d, J(4,5) = 9.5 Hz, 1H, 4-H), 3.41 (dd, J(5,6) = 7.1 Hz, 1H, 6-H), 3.31 - 3.27 (m, 2H, H-2,3), 2.97 (ddd, 1H, 5-H); C₆H₁₀O₄ (146.1): calcd C 49.31, H 6.90; found C 49.05, H 7.18.

1,5-Anhydro-2-deoxy-2-12,3:4,6-di-O-isopropylidene-5a-carba-*xylo***-hex-5(5a)enopyranosylaminoj-D-glucitol** (10) and **1,5-Anhydro-3-deoxy-3-12,3:4,6-di-O-isopropylidene-5a-carba-***xylo***-bex-5(5a)-enopyranosylaminoj-D-altritol** (11): A mixture of the epoxide 9 (29.0 mg, 0.198 mmol), valienamine 5 (39.0 mg, 0.153 mmol), and 2-propanol (0.5 mL) was heated in a sealed tube for 4 d at 120 °C, and then evaporated. TLC (butanone/toluene, 6:1, v/v) showed a formation of two products ($R_i = 0.31$ and 0.25). Chromatography of the products on a silica gel column (8 g)



with 2-butanone/toluene (3:1, v/v) as eluent gave 10 (35.7 mg, 58.2%) and 11 (15.5 mg, 25.3%) as a syrup.

Compound **10**, $[\alpha]_{D}^{23} = + 83.4$ (c = 0.87 in CHCl₃); ¹H NMR (270 MHz, CDCl₃, TMS): $\delta = 5.58$ (brd, J(1',5'a) = 4.4 Hz, 1H, 5'a-H), 4.50 (d, J(3',4') = 8.2 Hz, 1H, 4'-H), 4.46, 4.19 (2d, J(6'gem) = 14.3 Hz, each 1H, 6',6'-H). 4.08 (dd, $J(1_{eq}, 2) = 4.8$, J(1gem) = 11.0 Hz, 1H, 1_{eq} -H), 3.96 (dd, J(2',3') = 9.9 Hz, 1H, 3'-H), 3.87 (dd, J(5,6a) = 3.1, J(6gem) = 11.9 Hz. 1H, 6a-H), 3.78 (dd, J(5,6b) = 4.6, 1H, 6b-H), 3.73 (brdd, 1H, J(1',2') = 4.4, J(1',6') = 4.2 Hz, 1H, 1'-H), 3.56 (dd, 6'-H), 3.54 (dd, J(3,4) = 9.2, J(4,5) = 9.2 Hz, 1H, 4-H), 3.38 (dd, J(2,3) = 9.2 Hz, 1H, 3-H), 3.35-2.40 (m, 3H, 3OH), 3.26 (ddd, 1H, 5-H), 3.19 (dd, $J(1_{ac}, 2) = 11.0$ Hz, 1H, 1_{ac} -H), 2.80 (ddd, 1H, 2-H), 1.56, 1.49, 1.47, 1.43 (4s, each 3H, 2CMe₂); $C_{19}H_{31}$, NO₈ (401.5): cacld C 56.84, H 7.78, N, 3.49; found C 56.42, H, 8.28, N 3.43.

Compound 11, $[\alpha]_{D^3}^{D^3} = + 66.5$ (c = 0.34 in CHCl₃), ¹H NMR (270 MHz, CDCl₃, TMS): $\delta = 5.64$ (br d, J(1',5'a) = 3.3 Hz, 1 H, 5'a-H), 4.52 (d, 1 H, J(3',4') = 8.8 Hz, 1 H, 4'-H), 4.47, 4.21 (2 d, J(6'gem) = 14.3 Hz, each 1 H, 6',6'-H), 4.05-3.10 (m, 11 H, $1_{44}, 1_{eq}, 2.3, 4.5, 6, 1', 2', 3'$ -H), 1.56, 1.47, and 1.43 (3s, 3, 6, and 3 H, 2CMe₂); C₁₉H₃₁NO₈ (401.5): found C 56.45, H 8.24, N 3.47.

1,5-Anhydro-2-deoxy-2-{(1S,4R,5S,6S)-4,5,6-trihydroxy-3-hydroxymethylcyclohex-2-en-1-yljamino-D-glucitol (Salbostatin) (1), alternatively named 1,5-anhydro-2-deoxy-2-[5a-carba-xylo-hex-5(5a)-enopyranosyl]amino-D-glucitol: A mixture of 10 (15.7 mg, 0.039 mmol) and aqueous 70% acetic acid (1 mL) was stirred for 1 h at 60 °C, and then evaporated. Chromatography of the product on a column of Dowex 50 W-X2 (H⁺) (1 mL) with aqueous 1 M ammonia as eluent afforded 1 (12.2 mg, 97%) as a white solid, $[\alpha]_{D}^{25} = +130 (c = 1.20 \text{ in } H_2 \text{O}) [\text{ref. } [2]; [\alpha]_{D}^{20} = +115 (c = 1.20 \text{ in } H_2 \text{O}) [\alpha]_{D}^{20} = +115 (c = 1.20 \text{ in }$ in H₂O)], ¹H NMR (270 MHz, [D₆]DMSO/D₂O, 4:1, v/v, TMS): $\delta = 5.81$ (d, 1 H, J(1',2') = 3.3 Hz, 1 H, 2'-H), 4.05, 3.99 (2d, J(7'gem) = 14.4 Hz, each 1 H, 7',7'-H). 3.91 (dd, $J(1_{eq}, 2) = 4.6$, J(1 gem) = 11.2 Hz, 1 H, 1_{eq} -H), 3.79 (d, J(4', 5') = 5.5 Hz, 1H, 4'-H), 3.69 (d, $J(5,6a) \approx 0$, J(6 gem) = 12.1 Hz, 1H, 6a-H), 3.48 (dd, J(4',5') = 5.5, J(5',6') = 8.8 Hz, 1 H, 5'-H), 3.45 (dd, J(5,6b) = 5.9 Hz, 1 H, 6b-H), 3.41 (dd, J(1',6') = 4.8, J(5',6') = 8.8 Hz, 1 H, 6'-H), 3.23 (brdd, 1 H, 1'-H), 3.16-3.11 (m, 3H, 3,4,5-H), 3.02 (dd, $J(1_{a1},2) = 10.8$ Hz, 1_{a1} -H), 2.66 (ddd, 1H, 2-H); ¹³C NMR (67.5 MHz, [D₆]DMSO): δ = 140.38, 120.17, 81.67, 76.60, 72.90, 70.47, 70.15, 68.97, 61.37, 61.25, 57.54, 52.76.

3,4,6-Tri-O-acetyl-1,5-anhydro-2-deoxy-2-[(15,4R,55,65)-4,5,6-triacetoxy-3-acetoxymethylcyclohex-2-en-1-yl]amino-D-glucitol (12): Compound 10 (8.2 mg, 0.026 mmol) was treated with acetic anhydride (0.5 mL) in pyridine (1 mL) for 3 h at room temperature. The product was purified by chromatography on silica gel (1 g) with acetone/toluene (1:5, v/v) as eluent to yield 12 (15.4 mg, 98%) as a syrup. $[a]_{2}^{24} = + 66.6 (c = 0.75 \text{ in CHCl}_3), ^{1}\text{H NMR}$ (270 MHz, CDCl}_3, TMS): $\delta = 586$ (brd, J(1',2') = 4.8 Hz, 1 H, 2'-H), 5.53 (d, J(4',5') = 6.6 Hz, 1 H, 4'-H), 5.48 (dd, 1 H, J(5',6') = 9.7 Hz, 1 H, 5'-H), 4.97 (dd, 1 H, J(1',6') = 4.0 Hz, 1 H, 6'-H), 4.95 (dd, J(3,4) = 9.3, J(4,5) = 9.3 Hz, 1 H, 4'-H), 4.87 (dd, J(2,3) = 9.3 Hz, 1 H, 3-H), 4.64, 4.36 (2d, J(7'gem) = 13.2 Hz, each 1 H, 7',7'-H), 4.21 (dd, J(5,6a) = 4.8. J(6gem) = 12.5 Hz, 1 H, 6a-H), 4.09 (dd, 1 H, J(5,6b) = 2.2 Hz, 6b-H). 4.08 (dd, $J(1_{eq},2) = 4.8$, J(1gem) = 11.7 Hz, 1 H, 1_{eq} -H), 3.65 (brdd, J(1',2') = 2.2, J(1',6') = 4.0 Hz, 1 H, 1_{e1} -H), 2.88 (ddd, 1 H, 2-H), 2.10, 2.09, 2.06, 2.04, 2.03, 2.02 (6s, 3, 6, 3, 3, and 3 H, 7Ac); C_{27}H_{37}NO_{6} (471.6): calcd C 52.68, H 6.06, N 2.28; found C 53.08 H, 6.42, N, 2.37.

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